

## Colloidal characteristics in gradual binary associated systems of new “homogeneous” polyoxyethylene structured lipids

Călin Jianu<sup>1\*</sup>, Teodor Traşcă<sup>1</sup>, Adrian Riviş<sup>1</sup>, Adelina Jianu<sup>2</sup>, Corina Costescu<sup>1</sup>, Mihaela Cazacu<sup>1</sup>

<sup>1</sup>Faculty of Food Processing Technology, Banat's University of Agricultural Sciences and Veterinary Medicine “King Michael I of Romania” at Timișoara, Calea Aradului 119, RO-300645 Timișoara, Romania

<sup>2</sup>Faculty of Medicine, “Victor Babes” University of Medicine and Pharmacy Timisoara, 2nd Eftimie Murgu Square, RO-300041 Timisoara, Romania

Received: 27 September 2014; Accepted: 30 October 2014

### Abstract

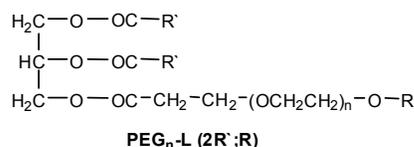
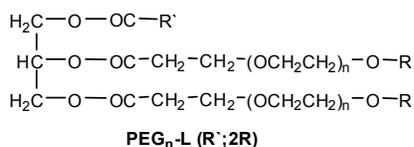
In series of structured lipids (SL) (I), conjugates PEG<sub>n</sub>-L (2R'; R) (R'; 2R), previously characterized by primary colloidal parameters: surface tensions (σ), critical micellar concentrations (CCM) and hydrophile/hydrophobe values (HLB) representatives with high lipophily (“colloidally disadvantaged”) as a result they have limited use in horticultural and agroalimentary processing due belonging to HLB = 1-8. The objective of the paper is to ameliorates in a controlled manner of this colloidal parameters through gradual binary monitorized association “homogeneous” (n = 9,18) polyoxyethylene (structured by Williamson procedure) conjugates PEG<sub>n</sub>-L (2R';1R) (R'; 2R) in series HLB<sub>1</sub>/HLB<sub>2</sub> = 4/10; HLB<sub>1</sub>/HLB<sub>2</sub> = 4/13; HLB<sub>1</sub>/HLB<sub>2</sub> = 6/10; HLB<sub>1</sub>/HLB<sub>2</sub> = 6/13 and ratios 10/90; 30/70; 50/50; 70/30; 90/10. Experimental results interpreted in correlation structures-HLB systems have imposed two new cumulated colloidal parameters: “homogeneous” cumulated oligomerisation degree (n<sub>CB</sub>), respectively hydrophile/hydrophobe cumulated value (HLB<sub>CB</sub>) “colloidally advantaged”. Performance of gradual binary association in this systems confirmed advantages and limits the premise for start-up and adopted work strategies.

**Keywords:** “homogeneous” polyoxyethylene conjugates, structured lipids, binary colloidal systems, cumulated colloidal competences

### 1. Introduction

The conjugates PEG<sub>n</sub>-L (2R';1R) (R';2R) (SL) (I) “colloidally disadvantaged” synthesized, purified and characterized in previous work [1,2], biodegradable biopolymers with highly lipophily, financially accessible and remarkable capacity of

derivatization and conditioning potential with adjustable solubility in typical environments to encapsulation, dispersion, processing allow the dosage limitation of the bioactive product introduced related to mass unit of the biopolymer [1,2,3-11].



We can present some of the major fields of other application for lipophyl structures:

- coated films for fresh horticultural products (storage, vegetables and fruits preservation). In this fields polyethyleneglycols (PEG<sub>n</sub>) and their fatty acids esters (E-430/431/433) can be accessed in agroalimentary processing receivers as emulsifiers, dispersing agents, binders or carriers by seclusion of active principles [12-16];
- wax coated films (E-907) have been elaborated to imitate the natural cover of fruits and vegetables. “Waxing” (cutin and suberine) (dewaxing) of fresh citric harvesting is done since XII<sup>th</sup> and XIII<sup>th</sup> century in China [17], but the pellicles limit the transfer of respiratory gases and induced fermentation. Gelatine pellicles have been proposed in the XIX<sup>th</sup> century and for the preservation of meat and other foods. Between 1930-1940 were sold as additivation products paraffin waxes which in melting were applied as a protector

to the fruits and vegetables, and afterwards were elaborated carnauba wax emulsions (E-903) [3,12,18,20,21]; to fight off degradation and simultaneously with the administration of fungicides [13,22,23,29]. Other additivation products include protectors against frosting, color additives, growth regulators or biological combatants [10,13,15-17,30-32] lipidic layer (monoglycerides acetates/E-472a associated with surfaceactive compounds) for the blocking of water transfer, fighting off “burnts”, MacIntosh apple decay and stains at Jonathan apples [10,11,17,19,33,34].

## 2. Materials and methods

### 2.1. Materials

- “homogeneous” polyoxyethylene conjugates PEG<sub>n</sub>-L (2R<sup>’</sup>;1R) (R<sup>’</sup>; 2R) (n = 3-9) with HLB<sub>1</sub> = 1-8, „colloidally disadvantaged”
- conjugates PEG<sub>n</sub>-L(2R<sup>’</sup>; 1R) (R<sup>’</sup>; 2R) with HLB<sub>2</sub> = 4-14 (Table 2) colloidally advantaged” [5];

**Table 1.** Physico-chemical (colloidal) parameters (HLB) of “colloidally disadvantaged” (HLB = 2,4-7,3) (selective list) conjugates series PEG<sub>n</sub>-L (2R<sup>’</sup>; 1R) (R<sup>’</sup>;2R) (n = 3, 6, 9) (R<sup>’</sup> = R<sup>’</sup><sub>s</sub>; R<sup>’</sup><sub>ca</sub>; R<sup>’</sup><sub>m</sub>; R<sup>’</sup><sub>co</sub>) R(NF; EH) accessed in evaluation HLB<sub>CB</sub> by binary systems gradually associated

No	Structure of conjugates PEG <sub>n</sub> -L (2R <sup>’</sup> ; 1R)	“Homogeneous” degree of oligomerisation	HLB <sub>1</sub> <sup>1</sup>
1	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>s</sub> ; 1 EH)	3	2,75
2	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>s</sub> ; 1NF)		2,54
3	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>ca</sub> ; 1 EH)		2,90
4	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>ca</sub> ; 1 NF)		2,43
5	PEG <sub>3</sub> -L (R <sup>’</sup> <sub>m</sub> ; 1 EH)		3,28
6	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>m</sub> ; 1 NF)		2,95
7	PEG <sub>3</sub> -L (R <sup>’</sup> <sub>co</sub> ; 1 EH)		2,93
8	PEG <sub>3</sub> -L (R <sup>’</sup> <sub>co</sub> ; 1 NF)		2,48
9	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>s</sub> ; 1 EH)	6	5,33
10	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>s</sub> ; 1 NF)		4,50
11	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>ca</sub> ; 1 EH)		5,39
12	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>ca</sub> ; 1 NF)		4,37
13	PEG <sub>3</sub> -L (R <sup>’</sup> <sub>m</sub> ; 1 EH)		5,70
14	PEG <sub>3</sub> -L (R <sup>’</sup> <sub>m</sub> ; 1 NF)		4,79
15	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>co</sub> ; 1 EH)		5,27
16	PEG <sub>3</sub> -L (2R <sup>’</sup> <sub>co</sub> ; 1 NF)		4,48
17	PEG <sub>9</sub> -L (2R <sup>’</sup> <sub>s</sub> ; 1 EH)	9	6,64
18	PEG <sub>9</sub> -L (R <sup>’</sup> <sub>s</sub> ; 1 NF)		6,32
19	PEG <sub>9</sub> -L (R <sup>’</sup> <sub>ca</sub> ; 1 EH)		6,64
20	PEG <sub>9</sub> -L (R <sup>’</sup> <sub>ca</sub> ; 1 NF)		6,25
21	PEG <sub>9</sub> -L (R <sup>’</sup> <sub>m</sub> ; 1 EH)		7,32
22	PEG <sub>9</sub> -L (R <sup>’</sup> <sub>m</sub> ; 1 NF)		6,74
23	PEG <sub>9</sub> -L (R <sup>’</sup> <sub>co</sub> ; 1 EH)		6,50
24	PEG <sub>9</sub> -L (R <sup>’</sup> <sub>co</sub> ; 1 NF)		6,35

**Table 2.** Conjugates PEG<sub>n</sub>-L (2R'; 1R) (R'; 2R) accessed in binary systems for optimization of “colloidally disadvantaged” structured lipids

No	Structure of association for some with hydrophile/hydrophobe values “colloidally disadvantaged” conjugates (HLB <sub>1</sub> ) and “colloidally advantaged” conjugates (HLB <sub>2</sub> )		
	HLB <sub>1</sub>		HLB <sub>2</sub>
1	4.00±0.5 PEG <sub>3</sub> -L(R'ca; 2NF) PEG <sub>3</sub> -L(R'ca; 2NF) PEG <sub>3</sub> -L(R'ca; 2NF)	10.00±0.5	PEG <sub>7</sub> -L(R'ca; 2EH) PEG <sub>7</sub> -L(R'co; 2EH) PEG <sub>7</sub> -L(R's; 1NF) PEG <sub>13</sub> -L(2R'm; 1NF) PEG <sub>13</sub> -L(2R'm; 1EH) PEG <sub>13</sub> -L(2R'co; 1EH)
		13.00±0.5	PEG <sub>13</sub> -L(R'ca; 2NF) PEG <sub>13</sub> -L(R's; 2NF) PEG <sub>13</sub> -L(R'm; 2NF) PEG <sub>13</sub> -L(R'co; 2NF) PEG <sub>13</sub> -L(R'ca; 2EH) PEG <sub>13</sub> -L(R's; 2EH) PEG <sub>13</sub> -L(R'co; 2EH)
2	6.00±0.5 PEG <sub>7</sub> -L(2R'ca; 1NF) PEG <sub>7</sub> -L(2R's; 1NF) PEG <sub>7</sub> -L(2R'co; 1NF) PEG <sub>7</sub> -L(2R'co; 1EH)	10.00±0.5	PEG <sub>13</sub> -L(R'ca; 2NF) PEG <sub>13</sub> -L(R'm; 2NF) PEG <sub>13</sub> -L(R's; 2NF) PEG <sub>13</sub> -L(R'co; 2NF) PEG <sub>13</sub> -L(R'ca; 2EH) PEG <sub>13</sub> -L(R's; 2EH) PEG <sub>13</sub> -L(R'co; 2EH)
		13.00±0.5	PEG <sub>13</sub> -L(R'ca; 2NF) PEG <sub>13</sub> -L(R's; 2NF) PEG <sub>13</sub> -L(R'm; 2NF) PEG <sub>13</sub> -L(R'co; 2NF) PEG <sub>13</sub> -L(R'ca; 2EH) PEG <sub>13</sub> -L(R's; 2EH) PEG <sub>13</sub> -L(R'co; 2EH)

propionic acid (Merck) (CAS 79-09-4); ethanol 98% (Merck) (CAS 64-17-5); phenol (Merck) (CAS 108-95-2).

**Table 3.** Dependence of cumulated “homogeneous” degrees of oligomerisation (n<sub>CB</sub>) for some conjugates PEG<sub>n</sub>-L(2R'; 1R) (R'; 2R) “colloidally disadvantaged” (n<sub>2</sub>) in various binary gradually associated systems

No	Binary systems		Binary gradually associated ratios				
	n <sub>1</sub>	n <sub>2</sub>	10/90	30/70	50/50	70/30	90/10
1	3	9	8,40	7,20	6,00	4,80	3,60
2	3	18	16,50	13,50	10,50	7,50	4,50
3	3	36	32,70	26,10	19,50	12,90	6,30
4	6	9	8,70	8,10	7,50	6,90	6,30
5	6	18	16,80	14,40	12,00	9,60	7,20
6	6	36	33,00	27,00	21,00	15,00	9,00
7	9	18	17,10	15,30	13,50	11,70	9,90
8	9	36	33,30	27,90	22,50	17,10	11,70
9	18	36	34,20	30,60	27,00	23,40	19,80

## 2.2. Methods

*Evaluation of water number (A) for polyoxyethylene chains (PEO) “homogeneous” (n = 3-18) per se and/or derivatized [1,41]:* Water number defined as the volume of aqueous phenol solution 2% added until the appearance a 1% solution of polyoxyethylene chains (PEO) “homogeneous” (n = 3-18) per se and/or derivatized chains, conjugates PEG<sub>n</sub>-L(2R';R), (R';2R), (ethyl alcohol solution 96% at 25°C), opalescence determined by the cleavage of

“hydrogen linkages” of the of polyoxyethylene chains (PEO) with water molecules in a process of double exchange (equilibrium) that exists the solution. It is accepted where:

$$HLB = 0.89A + 1.11$$

A = water number (mL solutions 2% phenol/ mL solution 10% of polyoxyethylene chains PEO in ethyl alcohol 96% at 25°C);

HLB = hydrophile/hydrophobe values “homogeneous” (n = 3-18) polyoxyethylene chains

(PEO), conjugates PEG<sub>n</sub>-L (2R'; R), (R'; 2R), studied.

Weigh 0,5 g conjugates PEG<sub>n</sub>-L(2R';R) (R';2R) to be studied in Jukov apparatus at 25 ± 0.5°C, dissolve in 5 mL ethyl alcohol 96 % and titrate it with aqueous phenol solutions 2 % from a biuret with double walls (at the same temperature as the Jukov apparatus).

Results are estimated with the relation:

$$A = \frac{\text{aqueous phenol solution 2 \% (mL)}}{\text{ethyl alcohol solution of PEG}_n\text{-L(2R'; R), (R'; 2R) (mL)}}$$

*Evaluation of HLB values after adapted Karabinos method [1,41]:* In 200-250 mL Erlenmeyer vessel weigh (analytical precision) 0.5 g conjugate PEG<sub>n</sub>-L(2R'; R), (R'; 2R), dissolve by stirring under controlled heating in 10-15 mL propionic acid (or 5-10 mL ethyl alcohol 98 % depending on the solubility of the studied product. Cooled by thermostatic bath at 25°C is titrated with aqueous phenol solution 2 % up to a permanent opalescence.

A is estimated with the relation:

$$A = \frac{\text{aqueous phenol solution 2 \% (mL)}}{\text{ethyl alcohol solution of PEG}_n\text{-L (2R'; R), (R'; 2R) (mL)}}$$

$$B = \frac{\text{aqueous phenol solution 2 \% (mL)}}{\text{propionic acid solution of PEG}_n\text{-L (2R'; R), (R'; 2R) (mL)}}$$

$$HLB = 0.89 (A)B + 1.11$$

### 3. Results. Discussions

In order to quantify as good as/possible the effects of the binary gradual association (n = 3, 6, 9) with that of the "colloidally disadvantaged" (HLB<sub>1</sub>=4-6) conjugates PEG<sub>n</sub>-L (2R';1R) (R';2R) and with the "colloidally advantaged" (HLB<sub>2</sub> = 10-13) (n<sub>2</sub> = 9, 18, 36) conjugates PEG<sub>n</sub>-L (2R';1R) (R';2R), there have been introduced in the paper two cumulated colloidal parameters [10,11].

- the cumulated "homogeneous" degree of oligomerisation by binary association (n<sub>CB</sub>) [24-26]:

$$n_{CB} = n_1 + n_2 \text{ [for PEG}_n\text{-L (R'; 2R)];}$$

$$n_{CB} = n_1 + n_2 = n_2 \text{ [for PEG}_n\text{-L (2R'; 1R)];}$$

$$\text{PEG}_3\text{-L (R'_{co}; 2NF); } n_{CB} = n_1 + n_2 = 3 + 3 = 6;$$

$$\text{PEG}_{18}\text{-L (2R'_{m}; 1EH); } n_{CB} = n_1 + n_2 = 0 + 18 = 18;$$

$$\text{PEG}_9\text{-L (R'_{co}; 2EH); } n_{CB} = n_1 + n_2 = 9 + 9 = 18;$$

$$\text{PEG}_{18}\text{-L (R'_{m}; 2EH); } n_{CB} = n_1 + n_2 = 18 + 18 = 36$$

- the hydrophile/hydrophobe cumulated value by gradual binary association (HLB<sub>CB</sub>) [27,28].

There have been selected two groups of "colloidally disadvantaged" conjugates PEG<sub>n</sub>-L (2R'; 1R) (R'; 2R) with the hydrophile/hydrophobe value (HLB<sub>1</sub> = 4.00 ± 0.5) belonging to the series PEG<sub>3</sub>-L(R'; 2NF) with three representants, respectively the series PEG<sub>9</sub>-L(2R';NF;EH) with the hydrophile/hydrophobe value HLB<sub>2</sub> = 6.00 ± 0.5 with four representants. In both groups of conjugates the "homogeneous" cumulated oligomerisation degree is n<sub>c</sub> = 6 and n<sub>c</sub> = 9 and the hydrocarbonated polyunsaturated chains la (R') belongs to the vegetal lipidic fractions, isolated, purified and characterized by the seeds (kernels) of the wildnut tree (R'<sub>ca</sub>) (*Aesculus hippocastanum*), grapes (R'<sub>s</sub>) (*Vitis vinifera*), coriander (R'<sub>co</sub>) (*Coriandri fructus*) and wiled rose (R'<sub>m</sub>) (*Rosa canina*) [1,6-9].

The conjugated structures selected belong to both series of ratios R'/R (11/2; 2/1) and to both types of hydro carbonated chains R(EH;NF).

We have also simultaneously decided to use two categories of "colloidally advantaged" conjugates PEG<sub>1</sub>-L (2R'; 1R) (R'; 2R) having the hydrophile/hydrophobe values HLB<sub>2</sub> = 10.00 ± 0.5 and belonging to the series PEG<sub>18</sub>-L (R'; 2EH/2NF) respectively to PEG<sub>18</sub>-L(2R'; EH/NF) (HLB<sub>2</sub> = 13.00 ± 0.5) with three representants conjugates series PEG<sub>18</sub>-L(2R'; EH) respectively ten representants.

In both cases of conjugates, the "homogeneous" cumulated oligomerisation degree is (n<sub>c</sub> = 18; n<sub>c</sub> = 36):

$$\text{PEG}_9\text{-L (R'; 2EH/2NF) with } n_c = 9 + 9 = 18;$$

$$\text{PEG}_{18}\text{-L (2R'; EH/NF) with } n_c = 9;$$

$$\text{PEG}_{18}\text{-L (R'; 2EH/2NF) with } n_c = 18 + 18 = 36$$

In order to optimize the HLB<sub>1</sub> "colloidally disadvantaged" values there has been done their gradual association in the series 10/90; 30/70; 50/50; 70/30; 90/10 with "colloidally advantaged" conjugates PEG<sub>n</sub>-L (2R';R) (R';2R) (HLB<sub>2</sub>). The consequences have been evaluated by correlation and by experimentally determining the values n<sub>CB</sub> (Table 3) and HLB<sub>CB</sub> (Tables 4, 5, 6).

**Table 4.** Dependence of cumulated “homogeneous” hydrophile/hydrophobe values for some conjugates PEG<sub>n</sub>-L(2R’; R) (R’; 2R) “colloidally disadvantaged” (HLB<sub>2</sub>) in various binary gradually associated systems

No	Binary systems		“Homogeneous” cumulated degree oligomerisation HLB <sub>CB</sub> in various binary gradually associated systems				
	HLB <sub>1</sub>	HLB <sub>2</sub>		HLB <sub>1</sub>	HLB <sub>2</sub>		HLB <sub>1</sub>
1	4	10	9,40	8,20	7	5,80	4,60
2	4	13	12,10	10,30	8,50	6,70	4,90
3	6	10	9,60	8,80	8,00	7,20	6,40
4	6	13	12,30	10,90	9,50	8,10	6,70

No	10/90 (HLB <sub>1</sub> =4)		30/70 (HLB <sub>1</sub> =4)		50/50 (HLB <sub>1</sub> =4)		70/30 (HLB <sub>1</sub> =4)		90/10 (HLB <sub>1</sub> =4)	
	HLB <sub>CB</sub>	HLB <sub>2</sub>								
Series 1	9,40	10	8,20	10	7,00	10	5,80	10	4,60	10
	12,10	13	10,30	13	8,50	13	6,70	13	4,90	13
	10/90 (HLB <sub>1</sub> =6)		30/70 (HLB <sub>1</sub> =6)		50/50 (HLB <sub>1</sub> =6)		70/30 (HLB <sub>1</sub> =6)		90/10 (HLB <sub>1</sub> =6)	
Series 2	HLB <sub>CB</sub>	HLB <sub>2</sub>								
	9,60	10	8,80	10	8,00	10	7,20	10	6,40	10
	12,30	13	10,90	13	9,50	13	8,10	13	6,70	13

No	10/90 (n <sub>1</sub> =3)		30/70 (n <sub>1</sub> =3)		50/50 (n <sub>1</sub> =3)		70/30 (n <sub>1</sub> =3)		90/10 (n <sub>1</sub> =3)	
	n <sub>CB</sub>	n <sub>2</sub>	n <sub>CB</sub>	n <sub>2</sub>	n <sub>CB</sub>	n <sub>2</sub>	n <sub>CB</sub>	n <sub>2</sub>	n <sub>CB</sub>	n <sub>2</sub>
Series 1	8,40	9	7,20	9	6,00	9	4,80	9	3,60	9
	16,50	18	13,50	18	10,50	18	7,50	18	4,50	18
	32,70	36	26,10	36	19,50	36	12,90	36	6,30	36
	8,40	9	7,20	9	6,00	9	4,80	9	3,60	9
Series 2	8,70	9	8,10	9	7,50	9	6,90	9	6,30	9
	16,80	18	14,40	18	12,00	18	9,60	18	7,20	18
	33,00	36	27,00	36	21,00	36	15,00	36	9,00	36
	n <sub>CB</sub>	n <sub>2</sub> (n <sub>1</sub> =9)	n <sub>CB</sub>	n <sub>2</sub> (n <sub>1</sub> =9)	n <sub>CB</sub>	n <sub>2</sub> (n <sub>1</sub> =9)	n <sub>CB</sub>	n <sub>2</sub> (n <sub>1</sub> =9)	n <sub>CB</sub>	n <sub>2</sub> (n <sub>1</sub> =9)
Series 3	17,10	18	15,30	18	13,50	18	11,70	18	9,90	18
	33,30	36	27,90	36	22,50	36	17,10	36	11,70	36
Limited series	n <sub>CB</sub>	n <sub>2</sub> (n <sub>1</sub> =18)								
	34,20	36								

**4. Conclusions**

The It is through that the major objectives of paper, to optimize in a controlled way the colloidal parameters of any “colloidally disadvantaged” conjugates PEG<sub>n</sub>-L (2R’;R) (R’;2R) through monitored gradual binary association (10/ 90; 30/70; 50/50; 70/30; 90/10) with “colloidally

advantaged” conjugates PEG<sub>n</sub>-L(2R’;R) (R;2R) (HLB<sub>2</sub>=4-14) is a benefit technological option. Experimental dates interpreted correlate with two new cumulated colloidal indicators (n<sub>CB</sub>; HLB<sub>CB</sub>) confirmed the premises for start-up and adopted work strategies.

**Table 5.** Dependence of binary “homogeneous” oligomerisation degrees cumulated n<sub>CB</sub> and binary cumulated hydrophile/hydrophobe values HLB<sub>CB</sub> in the series of conjugates PEG<sub>3</sub>-L(2R<sub>s</sub>’; EH) (R<sub>s</sub>’; 2EH)binary gradually associated (1-70 %) with “homogeneous” polyoxyethylene chains high oligomerised (n=30, 40, 50)

No	Conjugates	Colloidal parameter	Amount of “homogeneous” polyoxyethylene chain high oligomerised for binary graduall associated															
			EH <sub>30</sub>				EH <sub>40</sub>				EH <sub>50</sub>							
			1	10	20	50	70	1	10	20	50	70	1	10	20	50	70	
1	PEG <sub>3</sub> -L(2R <sub>s</sub> ’; EH)	n <sub>CB</sub>	3.27	5.70	8.40	16.50	21.90	3.37	6.70	10.40	21.50	28.90	3.47	7.70	12.40	26.50	35.90	
		HLB <sub>CB</sub> <sup>1)</sup>	2.90	4.28	5.82	10.42	13.49	2.90	4.29	5.83	10.46	13.54	2.91	4.36	5.97	10.81	14.04	
2	PEG <sub>3</sub> -L(R <sub>s</sub> ’; 2 EH) <sup>2,3)</sup>	n <sub>CB</sub>	6.24	8.40	10.80	18.00	22.80	6.34	9.40	12.80	23.00	29.80	6.44	10.40	14.80	28.00	36.80	
		HLB <sub>CB</sub> <sup>1)</sup>	5.41	6.61	7.88	11.71	14.26	5.46	6.61	7.90	11.75	14.31	5.47	6.69	8.04	12.10	14.82	

1) determined after Kamminos, J., (1955) [30], adapted Jianu C. Ph.D. 2006 [15];

2) cumulated “homogeneous” partial oligomerisation degree (n = 3);

3) cumulated hydrophile/hydrophobe values for conjugates “colloidally disadvantaged” PEG<sub>3</sub>-L(2R<sub>s</sub>’; EH) (HLB = 2.75); PEG<sub>3</sub>-L(R<sub>s</sub>’; 2EH) (HLB = 5.33)



**Compliance with Ethics Requirements.** Authors declare that they respect the journal's ethics requirements. Authors declare that they have no conflict of interest and all procedures involving human / or animal subjects (if exist) respect the specific regulation and standards.

#### References

- Jianu C., *Research concerning the potential of some metal ion complexing additives to improve food value in horticultural raw matter*. PhD thesis. 2006, Banat's University of Agricultural Sciences and Veterinary Medicine.
- Jianu C., Jianu I., Colloidal competences of new tailor-made lipids. *J Food Agric Environ.*, **2010**, 8, 148-155.
- Davidson R., *Handbook of Water-soluble Gums and Resins*, McGraw-Hill. New York, 1982
- Garret R., Smith N., Mehlschau J., *Apparatus and Method for Encapsulating Seeds and Like*, US Patent No. 4.806.357., 1989
- Govindram C.B., Krishnan V., Analysis of complex surfactant systems., *Tenside Surfactants Deterg.*, **1988**, 35(2), 104-107.
- Jianu, I., 1982, Romanian Patent, 80045.
- Jianu, I., 1983, Romanian Patent, 81853.
- Jianu, I., 1983, Romanian Patent, 81854.
- Drugarin C., Jianu I., The hydrolysis of  $\beta$ -[(p)-Alkylphenoxy polyethyleneoxy]-propionitrils under phase transfer catalysis - Part I: The synthesis of  $\beta$ -[(p)-Alkylphenoxy polyethyleneoxy]-propionitrils. *Tenside Detergents*, **1983**, 20, 128-129.
- Mark H., Bikales M, Overberger C., et al., *Encyclopedia of Polymer Science and Engineering*, Vol. 2, Wiley-Interscience, 1985, New York. 179-82.
- Mark H., Othmer D., Overberger C., et al., *Kirk-Othmer Encyclopedia of chemical Technology*, 3<sup>th</sup> edn. vol. 20. Wiley-Interscience, 1985, New-York. 219-20.
- Dadlani, M., Shenoy, V. and Seshu, D., Seed coating to improve stand establishment in rice, *Seed Sci. Technol.*, **1991**, 201, 307-13.
- Eckert M., Microencapsulated Flavours: Manufacture and Possible Applications Part 1, *ZFL.*, **1996**, 475, 67-70.
- Eckert M., Microencapsulated Flavours: Manufacture and Possible Applications Part 2, *ZFL.*, **1996**, 476, 63-65.
- Harris J., *Polyethyleneglycols Chemistry: Biotechnical and Biomedical Applications*, Harris. J.M. Ed., Plenum Press: New York, 1992
- Harris, J., Zaipey S., 1997. *Polyethyleneglycols Chemistry and Biological Applications*, Acs Symposium Series, 213<sup>th</sup> National Meeting of the American Chemical Society.
- Hardenburg, R., 1967, Wax and related coatings for horticultural products, A bibliography Agr. Res. Bull. No. 51-55. US Department of Agriculture, Washington. DC.
- Cottrell I., Kang K. and Kovacs P. 1980. Xanthan gum. In *Handbook of Water-soluble Gums and Resins*, ed. Davidson R.L., McGraw-Hill. New York. 1-30.
- Fisher, D. and Britton, J., 1940, Apple waxing experiments, *Sci. Agric.* 21:70-9.
- Garret, R., Mehlschau, J., Smith, N. et al., 1991, Gel encapsulation of tomato seeds, *Appl. Eng. Agric.* 71: 25-31.
- Kaplan, H., 1986, Washing. Waxing and adding in fresh Citrus Fruits, Eds. Wardowski, W.F., Nagy, S., and Grierson, W., AVI Publishing, Westport, CT. 379.
- Brown, E. 1984. Efficacy of citrus post-harvest fungicides applied in water or resin solution water wax. *Plant Dis.* 68:415-18.
- Miller, W.R., Chun, D., Risse, L.A., Hatton, T.T. and Hinsch T. 1988. Influence of selected fungicide treatments to control the development of decay in waxed or film wrapped Florida grapefruit, *Proc. 6<sup>th</sup> Int. Cong.*, 3:1471-7.
- Yousaf, T.I. 1994. Nuclear magnetic resonance (NMR) spectroscopy. In *Introduction to Surfactant Analysis*. Edited by Cullum, D.C.; Blackie Academic and Professional, Chapman and Hall: London; 297-317.
- ASTM-D-800-91. 1997. Standard Test Methods of Chemical Analysis of Industrial Metal Cleaning Compositions. American Society for Testing and Materials. Philadelphia. USA.
- Schmitt, T.M. 2002. *Analysis of surfactants*. 2<sup>nd</sup> Ed. Marcel Decker, Inc.; NY; 601-608.
- Weiß, J. 2001. *Ionen Chromatographie*. 3<sup>rd</sup> Ed. Wiley-VCH: Weinheim.
- Cullum, D.C. 1994. Basic techniques. In *Introduction to Surfactant Analysis*. Edited by Cullum, D.C.; Blackie Academic and Professional, Chapman and Hall: London; 45-48.
- Radnia, P. M. and Eckert J. W., 1988. Evaluations of imazalil efficacy in relation to fungicide formulation and wax formulation, *Proc. 6<sup>th</sup> Int. Cit. Cong.*, 3: 1427-34.

30. Baldwin, E. 1994. Edible coatings for fresh fruits and vegetables: past, present and future in *Edible Coatings and Films to improve Food Quality*, Eds. Krochata, J. Baldwin, E., and Nisperos-Carriedo, M., Technomic, Basel, Switzerland.
31. Harris, J., 1992. *Polyethyleneglycols Chemistry*, Plenum Press, New York.
32. Labuza, T. 1989. The effect of water activity on reaction kinetics of food deterioration. *Food Technology*, 34:36-41.
33. Orr, J., and Undertwood, T. 1987. Influence of gel coated seeds of germination time and tolerance of seedling tomatoes to postemergence herbicides, *Res. Prog. Rep. West Soc Wed. Sd. Western Society of Weed Science*. 130.
34. Warth, A. 1986. *The Chemistry and Technology of Waxes*, Reinhold New York. 37-192.
35. Jianu, C. 2014. Ethylene oxide homogeneous heterobifunctional acyclic oligomers. In *Oligomerization of Chemical and Biological Compounds*. Edited by Claire Lesieur, InTech: Croatia; 103.
36. Bujanca, G., Jianu, I. 2012. Synthesis of salified  $\beta$ -alkyl ( $C_{12}H_{25}/C_{18}H_{37}$ ) polyethyleneoxy ( $n=3-20$ ) propionamides, potential components in food sanitation (I). *J Food Agric Environ*, 10:182-188.
37. Jianu, I. 1984. *Aminoethers surfaceactive agents. Cationic surfaceactive agents*. PhD thesis. Polytechnic University.
38. Jianu, C. 2012. Colloidal competences of some food cleaning agents based on alkaline and ammonium nonionic soaps. *J Food Agric Environ*, 10:10-15.
39. Jianu, C., Trașcă T., Riviș, A., Miscă, C., Chiș, M., Jianu, I. 2009. Hydrolysis of  $\beta$ -alkyl ( $C_{12}H_{25}/C_{18}H_{37}$ ) polyethyleneoxy ( $n=3-20$ ) propionitriles in phase transfer catalysis conditions (II). *J Agroalim Proc Technol*, 15:79-87.
40. Jianu, C. 2012. Synthesis of nonionic-anionic colloidal systems based on alkaline and ammonium beta-nonylphenol polyethyleneoxy ( $n=3-20$ ) propionates/dodecylbenzenesulfonates with prospects for food hygiene. *Chem Cent J*, 6:95.
41. Karabinos, J., 1955, *Chem. Spec.*, 21:50.